Advances in the role of helper T cells in autoimmune diseases

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Abstract

Autoimmune diseases are primary immune diseases in which autoreactive antibodies or sensitized lymphocytes destroy and damage tissue and cellular components, resulting in tissue damage and organ dysfunction. Helper T cells may be involved in the pathogenesis of autoimmune diseases under certain conditions. This review summarizes recent research on the role of helper T cells in autoimmune diseases from two aspects, helper T cell-mediated production of autoantibodies by B cells and helper T cell-induced activation of abnormal lymphocytes, and provides ideas for the treatment of autoimmune diseases. The abnormal expression of helper T cells promotes the differentiation of B cells that produce autoantibodies, which leads to the development of different diseases. Among them, abnormal expression of Th2 cells and T follicular helper cells is more likely to cause antibody-mediated autoimmune diseases. In addition, abnormal activation of helper T cells also mediates autoimmune diseases, and a full understanding of their role in autoimmune diseases is helpful for providing ideas for the treatment of autoimmune diseases. Keywords: Helper T cells; B cells; Autoimmune disease

Introduction

Autoimmune diseases generally refer to immune effector cells such as cytotoxic T lymphocytes (CTLs), natural killer cells (NKs), macrophages, as well as immune effector molecules (complements, antibodies, cytokines, etc.) acting against their own tissues or cells to produce a pathological immune response, resulting in self-injury. A wide range of tissues can be damaged, including blood, skin, nerves, muscles, thyroid, bones and the gastrointestinal tract. Over 100 types of autoimmune diseases threaten people worldwide. At present, it is believed that individuals with certain genetic characteristics can develop autoimmune diseases due to the stimulation of some internal and external pathogenic factors or genetic mutation and modification of autoimmune response cells, resulting in abnormal activation of antigen presenting cells (APCs) or dendritic cells and an imbalance in the immunomodulatory network (the destruction of balance such as Th1/Th2 and Th17/regulatory T [Treg] cells) and changes in polyclonal activation or delayed apoptosis of autoimmune cells. There is also an autoimmune response in normal people, but autoimmune diseases occur only when the quantity or quality of the autoimmune response changes and the response intensity is strong enough to affect the function of tissues or cells. Many kinds of high titer autoantibodies or autoreactive sensitized lymphocytes may be detected in the blood of patients with autoimmune disease, which is

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known as autoimmune hyperimmunity. Autoantibodies and sensitized lymphocytes can be found in affected organs and tissues of some patients with autoimmune disease, and the extent of tissue damage depends on the distribution pattern of autoantigens targeted by autoantibodies or sensitized lymphocytes. This review describes the role of helper T cell-mediated autoantibodies produced by B cells and helper T cell-induced abnormal lymphocyte activation in the pathogenesis of autoimmune disease. According to the pathogenesis of different diseases, this paper provides ideas for the treatment of autoimmune disease in the future.

Autoimmune diseases mediated by B cells

Autoimmune hemolytic anemia (AIHA) is an acquired autoimmune disease that results in the production of autoantibodies against red blood cells (RBCs), causing shortened erythrocyte lifespan. However, the underlying mechanisms of antibody production are not fully understood. Studies of AIHA have found that the proliferation of T cells is enhanced *in vitro*.^[1] Recently, Th17 cells have been considered to be the key effector in AIHA development.^[2] The increase in Th17 cells and interleukin (IL)-17 secretion are closely related to the disease activity in AIHA patients. In an AIHA rat model, adoptive transfer of Th17 cells enhanced the response of anti-erythrocyte antibodies and increased the pathogenesis of AIHA, while neutralizing IL-17 *in vivo* eliminated the disease.^[3] Recently, a

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decrease in circulating CD4+ Treg cells and an increase in IL-10 and IL-12 levels were found in patients with AIHA.^[4] The imbalance in IL-10/IL-12 plays an important role in the pathogenesis or maintenance of AIHA. Amina et al^[5] found that CD25+ regulatory T cells controlled AIHA in C57BL/6J mice. Treatment with anti-CD25 antibody prior to immunization increased the incidence of AIHA to 90%, which may help to establish therapeutic strategies for the treatment of AIHA, as Treg cells are nonessential components of tolerance to the HEL-ovalbumin-Duffy (HOD) RBC autoantigen.^[6] Some data demonstrate the important role of T follicular helper (Tfh) cells in the control and induction of AIHA. Gao *et al*^[7] found that CD4+CXCR5+CD25+ Tfh cells were increased in an AIHA mouse model, suggesting that Tfh cells participate in B cell differentiation and anti-RBC antibody production. Strategies aimed at inhibiting Tfh development or function for curing AIHA should be emphasized.

Idiopathic thrombocytopenic purpura (ITP), which is similar to AIHA, is characterized by increased platelet destruction by autoantibodies directed against platelet glycoprotein. Autoreactive B lymphocytes secrete antiplatelet antibodies in ITP. The most common autoantibodies target platelet surface glycoprotein complexes GPIIb-IIIa and GPIb-IX after platelet internalization and degradation.^[8] Macrophages express platelet epitopes on their surface and secrete cytokines to stimulate CD4+ T cell clones,^[9] thus participating in B cell differentiation and antibody secretion. In patients with ITP, an increase in Th1 subsets leads to an imbalance in the Th1/Th2 subsets, which is conducive to the development of self-reactive B cells. Moreover, the cytokine polymorphism of Th1/Th2 also increases the susceptibility of ITP patients.^[10] Previous studies showed that Treg cells were significantly reduced during activity in ITP patients, and the levels and activity of Treg cells improved in patients who were treated with hormone or rituximab.^[11,12] In the peripheral blood of patients with ITP, the level of Treg cells was significantly decreased, while the level of Th17 cells was increased, leading to an imbalance in the Treg/Th17 ratio, which was also correlated with the disease activity in adults with ITP.^[13] In addition, Tfh cells may also be involved in the pathogenesis of ITP. Tfh cells provide IL-21-mediated auxiliary signals to B cells, promoting B cell proliferation and differentiation to plasma cells and the antibody response, and promote the differentiation and expansion of Th17 cells and Tfh cells.^[14] It has been reported that there is an increase in the proportion of circulating Tfh cells and splenic Tfh cells in ITP patients, particularly in anti-platelet antibody-positive patients.^[15] Th22 cells involved in the pathogenesis of ITP. Experiments by Zhan *et al*^[16]</sup> showed that Th22 cells are significantly increased in ITP patients and that Th22 cells are positively correlated with Th1 cells. Therefore, the unbalanced expression profile of Th22 cells in the peripheral blood is related to the pathogenesis of ITP.

Juvenile systemic sclerosis (jSSc) is a rare severe autoimmune disease with inflammatory mediators and autoantibodies that is similar to adult systemic sclerosis (SSc). Lymphocytes are the main cell type in jSSc lesions and mainly infiltrate the dermis and subdermis.^[17,18] By

stimulating fibroblasts to promote fibrosis, T cell activation and related cytokine release play key roles in the pathogenesis of SSc.^[19-21] Helper T cells and their effector cytokines have been found in skin biopsies,^[17,22,23] peripheral circulation and peripheral blood monocyte culture in patients with SSc.^[24-28] When the helper T cell subsets are out of balance, SSc develops. Previous studies have shown that Th2 cells and their related cytokines play a key role in adult SSc. A recent study by Mirizio *et al*^[27] analyzed 14 children with SSc and 24 healthy children. It was found that the proportion of circulating Th2 cells in children with SSc was significantly increased. Ten of these children were at the end of the disease; the level of Th17 cells in these children was significantly lower than that in the healthy control group. Th17 cells may contribute to inflammation in iSSc via production of associated proinflammatory cytokines in the earlier stage of disease. In addition, the levels of Th1-related cytokines (IL-1, tumor necrosis factor- α and interferon [IFN]- γ) increased in the peripheral blood of adult patients with SSc compared with those of the healthy control group, and their levels decreased over time. This means that the level of Th1 cells increases during the early stages of the disease.^[29,30] Similar to Th17 cells, Th1 cells may cause cellular inflammation in SSc by producing related proinflammatory cytokines during the early stages of the disease.^[27,31] Recently, Mirizio et al and Reiff et al^[27,32] reported the expression of Treg cells in iSSc patients functional Treg cells showed an overall downward trend compared with that of healthy controls and were positively correlated with clinical phenotype and the course of the disease. Part of the reason may be that Treg cells differentiate into Th17 cells and Th2 cells in blood and skin.^[33,34]

Myasthenia gravis (MG) is a chronic autoimmune disorder of neuromuscular transmission that results in muscle weakness. In MG, acetylcholine receptor (AChR)-specific T cells are important in inducing the production of pathogenic AChR antibodies. Abnormal activation of helper T cells is considered to be an important factor in the pathogenesis of MG. The levels of Th1 and Th17 cells in MG are increased, the related cytokines IFN-y and IL-17 are increased, and AChR-specific CD4+ T cells produce Th1 and Th17 cells in response to the acetylcholine receptor.^[35-38] The thymus plays a major role in the pathogenesis of MG with anti-AChR antibodies. Naturally occurring thymus-derived regulatory T cells are generated in the thymus and are key players in the suppression of the immune response. Several reports have illustrated the decreased number and function of thymic natural Treg (nTreg) cells in MG patients.^[39,40] This defect in nTreg cells attracts B cells and activates T cells, maintaining a chronic inflammatory state in the thymus.^[41] Imbalance in proinflammatory cytokines, Th1 and Th17 cells, and damage to Treg cells, as well as an imbalance in the immune response and inflammatory microenvironment are involved in the pathogenesis of MG. In patients with MG, Tfh cells play an important role in the selection and survival of B cells,^[42,43] the frequency of Tfh cells in the thymus of MG patients is increased, and the number of peripheral CXCR5+CD4+ T cells is also increased,[44] suggesting that Tfh cells are involved in the pathogenesis of MG. Further study in an experimental MG rat model

showed that in the early stage of experimental autoimmune MG, the number of pre-Tfh cells increased abnormally, which contributed to the formation of germinal centers and the secretion of antibodies by B cells. In the late stage of experimental autoimmune MG, Tfh cells showed AChR-specific induction, accompanied by the loss of pathological AChR in the muscle.^[45]

Systemic lupus erythematosus (SLE) is a chronic autoimmune disease that is characterized by autoantibodies against nuclear antigens, deposition of immune complexes and tissue damage to the kidney, skin, heart, and lung. A variety of immune cells and inflammatory mediators have been shown to be involved in the pathogenesis of SLE. CD4 + T cell dysfunction has been widely reported in patients with SLE.^[46] especially dysfunctional T and B cells. It has been found that the number of Tfh cells is increased in SLE patients, and Tfh cells can further differentiate into autoantibody-producing plasma cells by enhancing selfreactive B cell clones, which play a permanent role in the pathogenesis of SLE and eventually lead to autoimmunity.^[47] Some studies have reported that the decrease in Treg cells in the peripheral blood of patients with SLE is related to the disease activity index, immune abnormality and the type of tissue damage;^[48] Treg cells can be transformed into Th17-like cells in the development of SLE disease,^[49] which further suggests that these cells are involved in the activity of SLE. Th17 cell levels are significantly increased in the early stages of SLE compared with those of Treg cells and are associated with disease activity.^[50] SLE patients have higher Th17 levels, lower Treg levels, and a higher Th17/Treg ratio compared with those of the healthy control group, which are consistent with SLE activity.^[51]

T cell-mediated autoimmune disease

Multiple sclerosis (MS) is an autoimmune disease that specifically targets the white matter of the central nervous system and leads to demyelination. In MS and experimental autoimmune encephalomyelitis (EAE) mouse models, Th17 cells are the major promoter and participants in promoting pathology. Infusion of Th17 cells or injection of IL-17 effectively exacerbates the disease, while IL-17-deficient mice have alleviated pathology.^[52] In addition, studies have shown that the pathogenicity of Th17 cells in mice is related to the production of granulocytemacrophage colony-stimulating factor (GM-CSF).^[53] Therefore, downregulation of the immune response, especially Th17 cell differentiation and activation, may be an effective strategy for the treatment of MS. The levels of proinflammatory Th1 cytokines in patients with MS are higher than those in the control group, especially in the active stage of the disease. IFN- γ and GM-CSF have been detected in the cerebrospinal fluid and central nervous system of patients with MS. A strong Th1 response produces a high concentration of IFN-y-induced infiltration of important immune cells into the spinal cord, resulting in classical experimental autoimmune encephalomyelitis. Although the key role of Th1 cells in the initiation of MS and EAE has been identified, the mechanism of its involvement in the pathogenesis of demyelinating diseases has not been fully resolved. At present, the role and regulation of Th9 cells in MS patients

are still unclear. A study has shown that IL-9 levels in patients with MS were negatively correlated with inflammatory activity, neurodegeneration and disability, and high levels of IL-9 were associated with the loss of IL-17 in cerebrospinal fluid of patients with RRMS. IL-9 plays an immunomodulatory role in these patients.^[54] The role of IL-22 in MS has not been clarified, but it has been studied in recent years. In recent studies, there is evidence that Th22 cells are activated and serum IL-22 levels are increased in patients with MS, and Th22 cells are associated with Th17 cells, suggesting that Th22 and Th17 cells may play a synergistic role in the progression of MS.^[55]In Rolla *et al*^[56], the number of Th22 cells in peripheral blood and CSF increased in patients with recurrent and remission MS (RRMS), especially in the active stage of the disease. The expansion of Th22 cells is likely to help break through the blood-brain barrier, allowing for increased T cell infiltration, thus triggering MS disease. Recently, Perriard *et al*^[57] demonstrated the same results and found that serum IL-22 levels in patients with recurrent MS were significantly higher than those in healthy controls. This group also showed that astrocytes in the human brain express two subunits of IL-22 receptors. IL-22 colocalizes with these cells, which leads to the survivalpromoting characteristics of primary human astrocytes.

Th1 cells have been confirmed in psoriasis, and a large amount of data has shown that the number of Th1 cells that are capable of secreting IFN- γ is increased in the affected psoriatic skin. Th1-related cytokines such as IFN- γ and IL-2 are produced in the skin of most psoriatic lesions, which provides further evidence that Th1 cells cause psoriasis. However, in recent years, a number of studies have reported that other helper T cells are involved in the pathogenesis of psoriasis. Th17 cytokines, especially IL-17A, have been shown to play a key role in maintaining inflammation in psoriatic plaques. It has been observed that the number of CD4+ T cells producing IL-17 in psoriatic lesions is much higher than that in healthy skin. Activated Th17 cells enhance the inflammatory response of keratinocytes by forming a positive feedback loop around the IL-23/Th17 axis.^[58] The rapid and efficient results of anti-IL-17-based treatment strongly support the concept of IL-17 as a key amplification mechanism for determining the degree of skin manifestation in psoriasis.^[59] The association between genetic mutations in IL-23R, IL-12B and IL-23A and susceptibility to psoriasis strongly supports the key role of this axis in the pathogenesis of psoriasis.^[60] Eyerich *et al*^[61] reported that Th22 cells are mainly present in the skin of patients with psoriasis, and epidermal Th22 cells in psoriatic plaques still function after several years of disease remission, emphasizing the role of tissue-resident Th22 cells in the pathogenesis of psoriasis and the role of disease memory in the recurrence of psoriasis. Th9 cells were also detected in psoriatic lesions. In psoriatic lesions, the number of cells producing IL-9 was higher than that in healthy skin, and the expression of the IL-9 gene in psoriatic skin was significantly higher than that in normal skin of healthy subjects.^[62] Th9 cells enhance the ability of other T cell subsets to produce inflammatory cytokines and their presence in psoriatic lesions suggests that Th9 cells may also be involved in the initiation and maintenance of skin inflammation.^[63]

Aplastic anemia (AA) is a bone marrow failure syndrome in which hematopoietic stem cells are destroyed, leading to pancytopenia. At present, it is believed that T cell-mediated autoimmune imbalance is the main cause of acquired AA. Activated T cells induce apoptosis of hematopoietic stem cells. Oligoclonal amplification of cytotoxic CD8+ T cell imbalance has been confirmed in the bone marrow models of patients with AA in vitro. In vitro coculture of CD8+ T cells from untreated AA patients promotes apoptosis of normal CD3⁻ bone marrow cells and inhibit CD34⁺ cell colony formation.^[64] Damaged hematopoietic stem cells mature into self-reactive Th1 cells, which release IFN-y and tumor necrosis factor to transmit the cytotoxic cascade, killing and inhibiting other hematopoietic stem cells. In addition, an increase in Th17 cells was found in peripheral blood and bone marrow of patients with AA.^[65-67] Treg cells in bone marrow showed significant quantitative and quality defects,^[67] and the function of Treg cells in AA is impaired, as these cells cannot inhibit the autoreactivity of other T cell groups to normal tissues, including the bone marrow environment and hematopoi-etic stem cells,^[68,69] which ultimately leads to the failure of hematopoietic function.

Inflammatory bowel disease (IBD) is a group of complex diseases marked by chronic inflammation of the intestinal tract,^[70] including Crohn disease (CD) and ulcerative colitis (UC), and its specific etiology and pathogenesis have not been clarified. CD4+ T cells are considered to be the main driver of IBD, and CD4+ T cells are enriched in damaged tissues of patients with CD and UC; therefore, blocking or depleting CD4+ T cells is effective in patients with IBD. In IBD patients, CD has long been thought to be driven by Th1 cells, and the pathogenesis of UC has been associated with Th2 cells. In intestinal inflammation, IFN- γ binds to another Th1-related cytokine, tumor necrosis factor, to promote β-catenin signaling in intestinal epithelial cells, limiting their differentiation and proliferation.^[71] However, the role of IFN- γ in inflammatory bowel disease in mice is controversial. Powrie et al^[72] and Ito *et al*^[73] believe that IFN- γ promotes the development of the CD45RB^{hi}RAG adoptive transfer model and IBD DSS model disease. In these reports, a lack of IFN-y was associated with an overall reduction in the inflammatory response and tissue injury, as well as a reduction in other type 1-related chemokines and the ability to recruit other intestinal inflammatory cytokines. The number of regulatory T cells (CD4+CD8-CD25+) in inflammatory and noninflammatory tissues was higher than that in healthy controls.^[74] The ability of circulating Treg cells to inhibit autologous T cell proliferation decreased by approximately 60% in IBD patients compared with that of healthy controls,^[75] and circulating Treg cells are more likely to undergo apoptosis in inflammatory tissue.^[76] The expression of IL-17A^[77] and IL-17F^[78] increased in the intestinal tract of patients with IBD, and activated Th17 cells have been found in the intestinal mucosa and blood of patients with CD.^[79] In turn, these cells exacerbate inflammation by promoting the response of Th1 cells and Th17 cells. The increase in Th17-related cytokines may be due to the increase in lamina propria inflammation due to IL-17, IL-21 or IL-22 in Th17 cells, and the immune specificity of these cells is associated with the clinical activity of CD and ulcerative colitis.^[80] Recent studies have shown that Th9 cells and their cytokine IL-9 also promote IBD,^[81] and the transfer of Th9 cells leads to the exacerbation of UC in the intestinal mucosa of RAG-deficient mice, indicating that Th9 cells play a key role in the progression of IBD. In addition, the correlation between disease progression and IL-9 secreted by Th9 cells in patients with UC has also been recently confirmed.^[82,83] It has been found that IL-22 has a protective effect in an experimental model of colitis,^[84] and the number of Th22 cells producing IL-22 is reduced in patients with IBD.^[85] The deletion of Th22 cells in inflammatory mucosal cells of UC patients may lead to the upregulation of TGF- β .

In rheumatoid arthritis (RA), T cells and B cells may be involved in the pathogenesis to varving degrees, with T cell hyperactivity as the dominant immune response. The increase in Th17 cells in the peripheral blood and synovial fluid of patients with RA suggests the pathogenic role of Th17 cells in RA.^[86] Native T cells differentiate into Th17 cells through the participation of IL-1B, IL-6, IL-21, and TGF-B. IL-17 produced by Th17 cells induces fibroblast nuclear factor kappa B ligand receptor activator (RANKL) in the synovium to activate inflammation of a variety of immune cells and osteoclasts. In RA, there is an imbalance in Th17/Treg cells, and the activation degree of Th17 cells is significantly higher than that of Treg cells.^[87] In the synovial monocytes of patients with RA, the proportion of Th17 cells and the chemokine CCL20 is higher than that of the peripheral blood, while the proportion of Th1 cells is the opposite. Th17 cells of patients showed higher levels of RORyt and CCR6 expression compared with those of healthy subjects, especially in synovial fluid, which may lead to the selective migration of Th17 cells to inflammatory sites and lead to the development of rheumatoid arthritis. In recent years, the role of IL-22 in the pathogenesis and treatment of RA has become increasingly prominent.^[88] Th22 cells are subsets of CD4+ T cells that are characterized by the production of IL-22 but not IL-17 or IFN-y. IL-22 is the main characteristic cytokine of the Th22 subgroup. An increase in serum IL-22 is related to the disease activity of RA patients,^[89] promote osteoclast production and enhance bone destruction in arthritis mice, and the severity of the disease is significantly reduced in IL-22^{-/-}mice with collagen-induced arthritis.^[90] TGF- α is another important effective cytokine in Th22 cells and the main pathogenic factor of RA and it has a destructive effect on bone.

In conclusion, helper T cells play a very important role in the pathogenesis of autoimmune diseases. They not only participate in B cell-mediated autoimmune diseases but also promote B cells to differentiate into plasma cells and produce autoantibodies. Helper T cells also produce cytokines and chemokines that participate in the pathogenesis of diseases. Moreover, the ratio of helper T cell subsets can be out of balance through their own abnormal activation and decreased activity, resulting in disordered immune regulation, which leads to the occurrence of autoimmune diseases. Thus, helper T cells can be considered an effective therapeutic target for treating these disorders, and a full understanding of the changes in helper T cell subsets in autoimmune diseases is helpful to provide ideas for the treatment of autoimmune diseases.

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Conflicts of interest

None.

References

- 1. Fagiolo E. Immunological tolerance loss vs. erythrocyte self antigens and cytokine network disregulation in autoimmune hemolytic anaemia. Autoimm Rev 2004;3:53–59. doi: 10.1016/S1568-9972 (03)00085-5.
- Barcellini W, Clerici G, Montesano R, Taioli E, Morelati F, Rebulla P, et al. In vitro quantification of anti-red blood cell antibody production in idiopathic autoimmune haemolytic anaemia: effect of mitogen and cytoline stimulation. Br J Haematol 2000;111:452– 460. doi: 10.1046/j.1365-2141.2000.02380.x.
- 3. Xu L, Zhang T, Liu Z, Li Q, Xu Z, Ren T, *et al.* Critical role of Th17 cells in development of autoimmune hemolytic anemia. Exp. Hematol 2012;40:994–1004. doi: 10.1016/j.exphem.2012.08.008.
- Ahmad E, Elgohary T, Ibrahim H. Naturally occurring regulatory T Cells and interleukins 10 and 12 in the pathogenesis of idiopathic warm autoimmune hemolytic anemia. J Investig Allergol Clin Immunol 2011;21:297–304.
- Mqadmi A, Zheng X, Yazdanbakhsh K. CD4+CD25+ regulatory T cells control induction of autoimmune hemolytic anemia. Blood 2005;105:3746–3748. doi: 10.1182/blood-2004-12-4692.
- Richards AL, Kapp LM, Wang X, Howie HL, Hudson KE. Regulatory T Cells are dispensable for tolerance to RBC antigens. Front Immunol 2016;7:348. doi: 10.3389/fimmu.2016.00348.
- Gao Y, Jin H, Nan D, Yu W, Zhang J, Yang Y, *et al.* The role of T follicular helper cells and T follicular regulatory cells in the pathogenesis of autoimmune hemolytic anemia. Sci Rep 2019; 9:19767. doi: 10.1038/s41598-019-56365-3.
- Mcmillan R. Autoantibodies and autoantigens in chronic immune thrombocytopenic purpura. Semin. Hematol 2000;37:239–248. doi: 10.1016/s0037-1963(00)90102-1.
- Cines DB, Blanchette VS. Immune thrombocytopenic purpura. N Engl J Med 2002;346:995–1008. doi: 10.1056/NEJMra010501.
- Takahashi N, Saitoh T, Gotoh N, Nitta Y, Alkebsi L, Kasamatsu T, et al. The cytokine polymorphisms affecting Th1/Th2 increase the susceptibility to, and severity of, chronic ITP. BMC Immunol 2017;18:26. doi: 10.1186/s12865-017-0210-3.
- Zhang J, Zhang Q, Li Y, Tao L, Wu F, Shen Y, *et al.* Immune dysregulation in primary immune thrombocytopenia patients, hematology. Hematology 2018;23:1–7. doi: 10.1080/10245332. 2018.1435021.
- Zufferey A, Kapur R, Semple JW. Pathogenesis and therapeutic mechanisms in immune thrombocytopenia. J Clin Med 2017;6:16. doi: 10.3390/jcm6020016.
- Zhou ZJ, Zhang YS, Liang CX, Yang ZY. Role of th9, th17, treg cells levels and IL-9, IL-17 and TGF-beta expression in peripheral blood of patients withITP in pathogenesis of ITP (in Chinese). J Exp Hematol 2019;27:180–184. doi: 10.7534/j.issn.1009-2137.2019.01.029.
- Wu H, Deng Y, Zhao M, Zhang J, Zheng M, Chen G, et al. Molecular control of follicular helper T cell development and differentiation. Front Immunol 2018;9:2470. doi: 10.3389/ fimmu.2018.02470.
- 15. Yao X, Li C, Yang J, Wang G, Li C, Xia Y. Differences in frequency and regulation of T follicular helper cells between newly diagnosed and chronic pediatric immune thrombocytopenia. Blood Cells Mol Dis 2016;61:26–36. doi: 10.1016/j.bcmd.2016.06.006.
- Zhan FX, Li J, Fang M, Ding J, Wang Q. Importance of Th22 cell disequilibrium in immune thrombocytopenic purpura. Med Sci Monit 2018;24:8767–8772. doi: 10.12659/MSM.912528.
- Succaria F1, Kurban M, Kibbi AsG, Abbas O. Clinicopathological study of 81 cases of localized and systemic scleroderma. J Eur Acad Dermatol Venereol 2013;27:e191–e196. doi: 10.1111/j.1468-3083.2012.04581.
- Torres JE, Sánchez , Jorge L. Histopathologic differentiation, between localized and systemic scleroderma. Am J Dermatopathol 1998;20:242–245. doi: 10.1097/00000372-199806000-00003.

- Baraut J, Farge D, Jean-Louis F, Kesmandt H, Durant C, Verrecchia F, et al. Cytokines in systemic sclerosis. Pathol Biol 2012;60:127. doi: 10.1016/j.patbio.2009.11.003.
- Hasegawa M, Takehara K. Potential immunologic targets for treating fibrosis in systemic sclerosis: a review focused on leukocytes and cytokines. Semin Arthritis Rheum 2012;42:281–296. doi: 10.1016/j. semarthrit.2012.03.014.
- Fuschiotti P. T cells and cytokines in systemic sclerosis. Curr Opin Rheumatol 2018;30:594–599. doi: 10.1097/BOR.00000000 0000553.
- Keystone EC, Lau C, Gladman DD, Wilkinson S, Lee P, Shore A. Immunoregulatory T cell Subpopulations in patients with scleroderma using monoclonal antibodies. Clin Exp Immunol 1982;48:443– 448.
- Inoshita T, Whiteside TL, Rodnan GP, Taylor FH. Abnormalities of T lymphocyte subsets in patients with progressive systemic sclerosis (PSS, scleroderma). J Lab Clin Med 1981;97:264–277.
- 24. Kurasawa K, Hirose K, Sano H, Endo H, Shinkai H, Nawata Y, *et al.* Increased interleukin-17 production in patients with systemic sclerosis. Arthritis Rheum 2000;43:2455–2463. doi: 10.1002/ 1529-0131(200011)43:11<2455::AID-ANR12>3.0.CO;2-K.
- 25. Matsushita T, Hasegawa M, Hamaguchi Y, Takehara K, Sato S. Longitudinal analysis of serum cytokine concentrations in systemic sclerosis: association of interleukin 12 elevation with spontaneous regression of skin sclerosis. J Rheumatol 2006;33: 275–284.
- 26. Needleman BW, Wigley FM, Stair RW. Interleukin-1, interleukin-2, interleukin-4, interleukin-6, tumor necrosis factor alpha, and interferon-gamma levels in sera from patients with scleroderma. Arthritis Rheum 1992;35:67–72. doi: 10.1002/art.1780350111.
- Mirizio E, Marathi A, Hershey N, Ross C, Schollaert K, Salgado C, et al. Identifying the signature immune phenotypes present in pediatric localized scleroderma. J Invest Dermatol 2019;139:715– 718. doi: 10.1016/j.jid.2018.09.025.
- Valentini G1, Baroni A, Esposito K, Naclerio C, Buommino E, Farzati A, et al. Peripheral blood T lymphocytes from systemic sclerosis patients show both Th1 and Th2 activation. J Clin Immunol 2001;21:210–217. doi: 10.1023/a:1011024313525.
- 29. Gourh P, Arnett FC, Assassi S, Tan FK, Huang M, Diekman L, *et al.* Plasma cytokine profiles in systemic sclerosis: associations with autoantibody subsets and clinical manifestations. Arthritis Res Ther 2009;11. doi: 10.1186/ar2821.
- Dantas AT, Almeida AR, Sampaio MCPD, Cordeiro MF, Oliveira PSS, Mariz HA, *et al.* Different profile of cytokine production in patients with systemic sclerosis and association with clinical manifestations. Immunol Lett 2018;198:12–16. doi: 10.1016/j. imlet.2018.03.011.
- 31. Kurzinski KKC, Arkachaisri T, Feghali-Bostwick C, Torok KS. Circulating IP-10 and MCP-1 levels in active localized scleroderma. Arthritis Rheum 2011;63:S115.
- Reiff A, Weinberg KI, Triche T, Masinsin B, Mahadeo KM, Lin CH, et al. T lymphocyte abnormalities in juvenile systemic sclerosis patients. Clin Immunol 2013;149:146–155. doi: 10.1016/j. clim.2013.07.005.
- Slobodin G, Rimar D. Regulatory T cells in systemic sclerosis: a comprehensive review. Clin Rev Allergy Immunol 2017;52:1–8. doi: 10.1007/s12016-016-8563-6.
- MacDonald KG, Dawson NA, Huang Q, Dunne JV, Levings MK, Broady R. Regulatory T cells produce profibrotic cytokines in the skin of patients with systemic sclerosis. J Allergy Clin Immunol 2015;135:946–955.e9. doi: 10.1016/j.jaci 2014 12.1932.
- 35. Yi JS, Guidon A, Sparks S, Osborne R, Juel VC, Massey JM, et al. Characterization of CD4 and CD8 T cell responses in MuSK myasthenia gravis. J Autoimmun 2014;52:130–138. doi: 10.1016/j. jaut.2013.12.005.
- 36. Cao Y, Amezquita RA, Kleinstein SH, Stathopoulos P, Nowak RJ, O'Connor KC. Autoreactive T Cells from patients with myasthenia gravis are characterized by elevated IL-17, IFN-γ, and GM-CSF and diminished IL-10 production. J Immunol 2016;196:2075–2084. doi: 10 4049/jimmunol.1501339.
- Yilmaz V, Oflazer P, Aysal F, Parman YG, Direskeneli H, Deymeer F, *et al.* B cells produce less IL-10, IL-6 and TNF-α in myasthenia gravis. Autoimmunity 2015;48:201–207. doi: 10.3109/08916934.2014. 992517.
- Villegas JA, Van Wassenhove J, Le Panse R, Berrih-Aknin S, Dragin N. An imbalance between regulatory T cells and T helper 17 cells in

acetylcholine receptor-positive myasthenia gravis patients. Ann N Y Acad Sci 2018;1413:154–162. doi: 10.1111/nyas.13591.

- Gradolatto A, Nazzal D, Truffault F, Bismuth J, Fadel E, Foti M, *et al.* Both Treg cells and Tconv cells are defective in the Myasthenia gravis thymus: roles of IL-17 and TNF-α. J Autoimmun 2014;52:53–63. doi: 10 1016/j jaut 2013 12.015.
- Nishimura T, Ínaba Y, Nakazawa Y, Omata T, Akasaka M, Shirai I, et al. Reduction in peripheral regulatory T cell population in childhood ocular type myasthenia gravis. Brain Dev 2015;37:808– 816. doi: 10.1016/j.braindev.2014.12.007.
- 41. LePanse R, Cizeron-Clairac G, Cuvelier M, Truffault F, Bismuth J, Nancy P, et al. Regulatory and pathogenic mechanisms in human autoimmune myasthenia gravis. Ann N Y Acad Sci 2008;1132:135– 142. doi: 10.1196/annals.1405.019.
- 42. Song Y, Zhou L, Miao F, Chen G, Zhu Y, Gao X, et al. Increased frequency of thymic T follicular helper cells in myasthenia gravis patients with thymoma. J Thorac Dis 2016;8:314–322. doi: 10.21037/jtd.2016.03.03.
- 43. Zhang X, Liu S, Chang T, Xu J, Zhang C, Tian F, *et al.* Intrathymic Tfh/B cells interaction leads to ectopic GCs formation and anti-AChR antibody production: central role in triggering MG occurrence. Mol Neurobiol 2016;53:120–131. doi: 10.1007/s12035-014-8985-1.
- 44. Wen Y, Yang B, Lu J, Zhang J, Yang H, Li J. Imbalance of circulating CD4(+)CXCR5(+)FOXP3(+) Tfr-like cells and CD4(+)CXCR5(+) FOXP3(-) Tfh-like cells in myasthenia gravis. Neurosci Lett 2016;630:176–182. doi: 10.1016/j.neulet.2016.07.049.
- 45. Cui YZ, Qu SY, Chang LL, Zhao JR, Mu L, Sun B, et al. Enhancement of T follicular helper cell-mediated humoral immunity reponses during development of experimental autoimmune myasthenia gravis. Neurosci Bull 2019;35:507–518. doi: 10.1007/s12264-019-00344-1.
- Konya C, Paz Z, Tsokos GC. The role of T cells in systemic lupus erythematosus: an update. Curr Opin Rheumatol 2014;26:493–501. doi: 10.1097/BOR.00000000000082.
- Zhang X, Lindwall E, Gauthier C, Lyman J, Spencer N, Alarakhia A, et al. Circulating CXCR5+CD4+helper T cells in systemic lupus erythematosus patients share phenotypic properties with germinal center follicular helper T cells and promote antibody production. Lupus 2015;24:909–917. doi: 10.1177/0961203314567750.
 Yang X, Wang W, Xu J, Zhang MS, Mei H, Shen Y, et al. Significant
- Yang X, Wang W, Xu J, Zhang MS, Mei H, Shen Y, *et al.* Significant association of CD4+CD25+Foxp3+ regulatory T cells with clinical findings in patients with systemic lupus erythematosus. Ann Transl Med 2019;7:93. doi: 10.21037/atm.2019.01.38.
- 49. Jiang C, Wang H, Xue M, Lin L, Wang J, Cai G, *et al.* Reprograming of peripheral Foxp3+ regulatory T cell towards Th17-like cell in patients with active systemic lupus erythematosus. Clin Immunol 2019;209:108267. doi: 10.1016/j.clim.2019.108267.
- Talaat RM, Mohamed SF, Bassyouni IH, Raouf AA. Th1/Th2/Th17/ Treg cytokine imbalance in systemic lupus erythematosus (SLE) patients: Correlation with disease activity. Cytokine 2015;72:146– 153. doi: 10.1016/j.cyto.2014.12.027.
- Yuliasih Y, Rahmawati LD, Putri RM. Th17/Treg ratio and disease activity in systemic lupus erythematosus. Caspian J Intern Med 2019;10:65–72. doi: 10.22088/cjim.10.1.65.
- Komiyama Y, Nakae S, Matsuki T, Nambu A, Ishigame H, Kakuta S, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. J Immunol 2006; 177:566–573. doi: 10.4049/jimmunol.177.1.566.
- 53. Nacka-Aleksic M, Djikic J, Pilipovic I, Stojic-Vukanic Z, Kosec D, Bufan B, *et al.* Male rats develop more severe experimental autoimmune encephalomyelitis than female rats: sexual dimorphism and diergism at the spinal cord level. Brain Behav Immun 2015;49:101–118. doi: 10.1016/j.bbi.2015.04.017.
- Ruocco G, Rossi S, Motta C, Macchiarulo G, Barbieri F, De Bardi M, et al. T helper 9 cells induced by plasmacytoid dendritic cells regulate interleukin-17 in multiple sclerosis. Clin Sci (Lond) 2015;129:291– 303. doi: 10.1042/CS20140608.
- 55. Xu W, Li R, Dai Y, Wu A, Wang H, Cheng C, et al. IL-22 secreting CD4+ T cells in the patients with neuromyelitis optica and multiple sclerosis. J Neuroimmunol 2013;261:87–91. doi: 10.1016/j.jneuroim.2013.04.021.
- 56. Rolla S, Bardina V, De Mercanti S, Quaglino P, De Palma R, Gned D, et al. Th22 cells are expanded in multiple sclerosis and are resistant to IFN-β. J Leukoc Biol 2014;96:1155–1164. doi: 10.1189/jlb.5A0813-463RR.

- 57. Perriard G, Mathias A, Enz L, Canales M, Schluep M, Gentner M, *et al.* Interleukin-22 is increased in multiple sclerosis patients and targets astrocytes. J Neuroinflammation 2015;12:119. doi: 10.1186/s12974-015-0335-3.
- Lowes MA, Russell CB, Martin DA, Towne JE, Krueger JG. The IL-23/T17 pathogenic axis in psoriasis is amplified by keratinocyte responses. Trends Immunol 2013;34:174–181. doi: 10.1016/j. it.2012.11.005.
- 59. Cho JH, Feldman M. Heterogeneity of autoimmune diseases: pathophysiologic insights from genetics and implications for new therapies. Nat Med 2015;21:730–738. doi: 10.1038/nm.3897.
- Harden Jamie L, Krueger James G, Bowcock Anne M. The immunogenetics of Psoriasis: a comprehensive review. J Autoimmun 2015;64:66–73. doi: 10.1016/j.jaut.2015.07.008.
- Cheuk S, Wikén M, Blomqvist L, Nylén S, Talme T, Ståhle M, et al. Epidermal Th22 and Tc17 cells form a localized disease memory in clinically healed psoriasis. J Immunol 2014;192:3111–3120. doi: 10.4049/jimmunol.1302313.
- Sgambelluri F, Diani M, Altomare A, Frigerio E, Drago L, Granucci F, *et al.* A role for CCR5(+)CD4 T cells in cutaneous psoriasis and for CD103(+) CCR4(+) CD8 Teff cells in the associated systemic inflammation. J Autoimmun 2016;70:80–90. doi: 10.1016/j. jaut.2016.03.019.
- 63. Schlapbach C, Gehad A, Yang C, Watanabe R, Guenova E, Teague JE, *et al.* Human TH9 cells are skin-tropic and have autocrine and paracrine proinflammatory capacity. Sci Transl Med 2014;6:219ra8. doi: 10.1126/scitranslmed.3007828.
- 64. Nakao S, Takami A, Takamatsu H, Zeng W, Sugimori N, Yamazaki H, *et al.* Isolation of a T-cell clone showing HLA-DRB1*0405-restricted cytotoxicity for hematopoietic cells in a patient with aplastic anemia. Blood 1997;89:3691–3699.
- Luzzatto L, Risitano AM. Advances in understanding the pathogenesis of acquired aplastic anaemia. Br J Haematol 2018;182:758–776. doi: 10.1111/bjh.15443.
- 66. Gonzaga VF, Wenceslau CV, Lisboa GS, Frare EO, Kerkis I. Mesenchymal stem cell benefits observed in bone marrow failure and acquired aplastic anemia. Stem Cells Int 2017;2017:8076529. doi: 10.1155/2017/8076529.
- Shallis RM, Ahmad R, Zeidan AM. Aplastic anemia: etiology, molecular pathogenesis and emerging concepts. Eur J Haematol 2018;101:711–720. doi: 10.1111/ejh.13153.
- Kordasti S, Marsh J, Al-Khan S, Jiang J, Smith A, Mohamedali A, et al. Functional characterization of CD4+ T cells in aplastic anemia. Blood 2012;119:2033–2043. doi: 10.1182/blood-2011-08-368308.
- 69. Kordasti S, Costantini B, Seidl T, Perez Abellan P, Martinez Llordella M, McLornan D, *et al.* Deep phenotyping of Tregs identifies an immune signature for idiopathic aplastic anemia and predicts response to treatment. Blood 2016;128:1193–1205. doi: 10.1182/ blood-2016-03-703702.
- Weigmann B, Neurath MF. Th9 cells in inflammatory bowel diseases. Semin Immunopathol 2017;39:89–95. doi: 10.1007/s00281-016-0603-z.
- Nava P, Koch S, Laukoetter MG, Lee WY, Kolegraff K, Capaldo CT, et al. Interferon-γ regulates intestinal epithelial homeostasis through converging β-catenin signaling pathways. Immunity 2010;32:392– 402. doi: 10.1016/j.immuni.2010.03.001.
- 72. Powrie F, Leach MW, Mauze S, Menon S, Caddle LB, Coffman RL. Inhibition of Th1 responses prevents inflammatory bowel disease in scid mice reconstituted with CD45RBhi CD4+ T cells. Immunity 1994;1:553–562.
- 73. Ito R, Shin-Ya M, Kishida T, Urano A, Takada R, Sakagami J, et al. Interferon-gamma is causatively involved in experimental inflammatory bowel disease in mice. Clin Exp Immunol 2006;146:330–338. doi: 10.1111/j.1365-2249.2006.03214.x.
- 74. Saruta M, Yu QT, Fleshner PR, Mantel PY, Schmidt-Weber CB, Banham AH, *et al.* Characterization of FOXP3+CD4+ regulatory T cells in Crohn's disease. Clin Immunol 2007;125:281–290. doi: 10.1016/j.clim.2007.08.003.
- 75. Ueno A, Jijon H, Chan R, Ford K, Hirota C, Kaplan GG, et al. Increased prevalence of circulating novel IL-17 secreting Foxp3 expressing CD4+ T cells and defective suppressive function of circulating Foxp3+ regulatory cells support plasticity between Th17 and regulatory T cells in inflammatory bowel disease patients. Inflamm Bowel Dis 2013;19:2522–2534. doi: 10.1097/MIB.0b013e3182a85709.

- 76. Veltkamp C, Anstaett M, Wahl K, Möller S, Gangl S, Bachmann O, *et al.* Apoptosis of regulatory T lymphocytes is increased in chronic inflammatory bowel disease and reversed by anti-TNFα treatment. Gut 2011;60:1345–1353. doi: 10.1136/gut.2010.217117.
- 77. Fujino S, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, et al. Increased expression of interleukin 17 in inflammatory bowel disease. Gut 2003;52:65–70. doi: 10.1136/gut.52.1.65.
- Geremia A, Arancibia-Cárcamo CV, Fleming MP, Rust N, Singh B, Mortensen NJ, et al. IL-23-responsive innate lymphoid cells are increased in inflammatory bowel disease. J Exp Med 2011;208:1127– 1133. doi: 10.1084/jem.20101712.
- Kleinschek MA, Boniface K, Sadekova S, Grein J, Murphy EE, Turner SP, et al. Circulating and Gut-resident Human Th17 cells express CD161 and promote intestinal inflammation. J Exp Med 2009; 206:525–534. doi: 10.1084/jem.20081712.
- Jiang W, Su J, Zhang X, Cheng X, Zhou J, Shi R, *et al.* Elevated levels of Th17 cells and Th17-related cytokines are associated with disease activity in patients with inflammatory bowel disease. Inflamm Res 2014;63:943–950. doi: 10.1007/s00011-014-0768-7.
- Ye Y, Pang Z, Chen W, Ju S, Zhou C. The epidemiology and risk factors of inflammatory bowel disease Int J Clin Exp Med 2015; 8:22529–22542.
- 82. Gerlach K, Hwang Y, Nikolaev A, Atreya R, Dornhoff H, Steiner S, et al. TH9 cells that express the transcription factor PU.1 drive T cellmediated colitis via IL-9 receptor signaling in intestinal epithelial cells. Nat Immunol 2014;15:676–686. doi: 10.1038/ni.2920.
- Nalleweg N, Chiriac MT, Podstawa E, Lehmann C, Rau TT, Atreya R, *et al.* IL-9 and its receptor are predominantly involved in the pathogenesis of UC. Gut 2015;64:743–755. doi: 10.1136/gutjnl-2013-305947.

- 84. Dudakov JA, Hanash AM, van den Brink MR. Interleukin-22: immunobiology and pathology. Annu Rev Immunol 2015;33:747– 785. doi: 10.1146/annurev-immunol-032414-112123.
- Leung JM, Davenport M, Wolff MJ, Wiens KE, Abidi WM, Poles MA, et al. IL-22-producing CD4+ cells are depleted in actively inflamed colitis tissue. Mucosal Immunol 2014;7:124–133. doi: 10.1038/mi.2013.31.
- 86. Leipe J, Schramm MA, Prots I, Schulze-Koops H, Skapenko A. Increased Th17 cell frequency and poor clinical outcome in rheumatoid arthritis are associated with a genetic variant in the IL4R gene, rs1805010. Arthritis Rheumatol 2014;66:1165–1175. doi: 10.1002/art.38343.
- Takayanagi H. Osteoimmunology and the effects of the immune system on bone. Nat Rev Rheumatol 2009;5:667–676. doi: 10.1038/ nrrheum.2009.217.
- Xie Q, Huang C, Li J. Interleukin-22 and rheumatoid arthritis: emerging role in pathogenesis and therapy. Autoimmunity 2015;48:69–72. doi: 10.3109/08916934.2014.959165.
- 89. Da Rocha LF Jr, Duarte ÂL, Dantas AT, Mariz HA, Pitta Ida R, Galdino SL, *et al.* Increased serum interleukin 22 in patients with rheumatoid arthritis and correlation with disease activity. J Rheumatol 2012;39:1320–1325. doi: 10.3899/jrheum.111027.
- Corneth OB, Reijmers RM, Mus AM, Asmawidjaja PS, van Hamburg JP, Papazian N, *et al.* Loss of IL-22 inhibits autoantibody formation in collagen-induced arthritis in mice. Eur J Immunol 2016;46:1404–1414. doi: 10.1002/eji.201546241.

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